Can lactoferrin modulate the immunostimulant activity of levamisole in rats immunosuppressed by cyclophosphamide?

Laktoferrin siklofosfamid ile immunsuprese ratlarda levamizol’un immün stimulan etkisini düzenleyebilir mi?

Wafaa Abdou Mohamed Mohamed

ABSTRACT

Objective: The aim of this study was to study the immunomodulatory activity improvement of levamisole by using lactoferrin when applied to immunosuppressed rat model.

Methods: The study was designed as follows, 140 male albino rats (250-280 g) 14 weeks old were used in our work. Rats were randomly divided into seven groups, 20 in each. The group I was kept as a control, group II was given cyclophosphamide (CYP) at a single intraperitoneal dose of (250 mg/kg body weight), group III CYP and lactoferrin (Lac) treated group, group IV orally administrated Lac only (0.5%) in drinking water, group V treated with CYP and levamisole, group VI administrated levamisole orally at a dose of (2.5 mg/kg body weight) and group VII was given CYP, Lac and levamisole. Animals were sacrificed and two separate blood samples were collected after 21 days from the beginning of the experiment for measuring the total and differential leukocyte count, serum total proteins, albumin, alpha globulin, beta globulin and gamma globulin, Nitric oxide (NO) production and lysozyme activity.

Results: CYP group showed significant decrease in the above mentioned parameters, which were improved after administration of both lactoferrin and levamisole.


Key words: Bovine lactoferrin (Lac), Levamisole , Cyclophosphamide(CYP), Immunomodulation, Lysozyme assay

INTRODUCTION

Immunostimulants are agents that trigger the non-specific immune response and result in enhanced disease resistance. Several compounds have been reported to have immunostimulation properties. Many of these are derivatives or cellular components of bacterial, fungal or animal origin such as laminarin, barley, glucan, lactoferrin, levamisole, lipopolysaccharides, curdian, scleroglucan, zymosan, inulin, chitosan, glucans, dextran, lentinant, saponins, herbal extracts, peptidoglycans [1].

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Cyclophosphamide (CYP) is an alkylating agent widely used as cancer chemotherapy and autoimmune disease therapy [2]. CYP is a well-known and powerful immunosuppressive agent with dose-specific effects. Low dose of CYP acts mainly on B cell regions in lymphoid organs, but high doses have an effect upon both B and T cell regions [3]. Levamisole is mainly used as an anthelmintic agent in veterinary purpose [4] but in some countries, its use is limited to immunomodulatory agent in humans in some cancers. It is having an immunostimulating effect in immunosuppressed condition [5] but it has been found that it is also having a useful effect in autoimmune diseases like nephrotic syndrome and rheumatoid arthritis. It helps to make steroid free period of up to 6 months to 1 year in nephrotic syndrome [6,7].

Lactoferrin (Lac) is an 80 kDa multifunctional glycoprotein belonging to the transferrin family. Lac is primarily present in milk, and is also found in other biological fluids, such as saliva, tears, bile and pancreatic juice [8]. It has been widely documented that Lac displays antimicrobial activity against many different pathogenic agents. This activity was attributed to its ability, to bind iron with a high affinity and unlike transferrin, retain its bound iron under acidic conditions. Lac is considered to be a part of the innate immune system and takes part in specific immune reactions, but in an indirect way [9]. Lac has a wide range of effects on the immune system, both in vivo and in vitro [10]. Lactoferrin is a cell-secreted mediator that bridges innate and adaptive immune function in mammals. It is a pleiotropic molecule that directly assists in the influence of presenting cells for the development of T-helper cell polarization [11].

The objective of the present research work is to check the effect of to study the improvement of the levamisole immunomodulation action when given with lactoferrin.

METHODS

Animals

140 male albino rats (250-280 g) 14 weeks old were obtained from the faculty of Veterinary Medicine, Zagazig University to use for the study. The rats were distributed in seven groups, housed in solid-bottomed cages containing bedding of wood shavings and were allowed food and water ad libitum. The room temperature was maintained at 21-24°C, and a 12 h light/dark cycle was employed. All animals were acclimatized for 1 week before starting the experiment. The ethical standards guidelines for the care and use of laboratory animals provided by the animal ethics committee (institutional or national) and with the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) were used.

Chemicals

Immunostimulant agent (levamisole 10%) levamisole hydrochloride (pharma sewede, Egypt). Bovine lactoferrin was purchased from Sigma-Aldrich Chemie GmbH. Endoxan® injection vial of Endoxan 1 g contains: 1.069 g cyclophosphamide monohydrate (equivalent to 1 g anhydrous cycophosphamide) as the active ingredient was manufactured by industri-as Farmaceuticas Almirall Prodesfarma S.L.C/San Juan no.9.08560 Manlleu/Spain for Baxter Oncology GmbH Kantstrasse 2 D-33790 Halle, Germany. All the other chemicals were purchased from standard local sources. All other reagents used were of analytical grade.

Experimental design

Rats were randomly divided into seven groups; each is consisting of twenty rats. The groups treated as follows. The group I was kept as control, group II was given CYP at a single intraperitoneal dose of (200 mg/kg body weight) on the first day of the experiment according to [12], group III was administered CYP and Lac, group IV orally administrated Lac only (0.5%) in drinking water during the 21 days of the experiment according to [13], Group V treated with CYP and levamisole. Group VI orally administrated levamisole at a dose of (2.5 mg/kg body weight) according to [14]. Group VII was treated with CYP, Lac and levamisole.

Sampling

Animals were anesthetized with diethyl ether then were scarificed at the end of the experiment, two separate blood samples were collected from each of five rats from each group; the 1st sample was taken in EDTA tubes for measurement of total and differential leukocytes count according to [15,16], 2nd sample was taken without anticoagulant and kept 30 minutes at room temperature then centrifuged at 3000 rpm for 10 minutes and the clear serum was separated carefully and storage at -20°C for the measurement of some immunological parameters.

Evaluation of some immunological parameters

Serum total protein level was determined according to Henry et al [17], the serum albumin level was determined by the method of Doumas et al [18]
using the kits provided by Diamond Diagnostics. Regarding to immunological studies, Immunoelectrophoresis of serum proteins has been done using cellulose acetate according to Henry et al [17], lysozyme activity in blood plasma was determined by the turbidimetric method of [19] modified by [20] and nitric oxide production assay was performed as mentioned by [21].

Lysozyme activity

Whole blood samples were centrifuged for 5 min at 1000 g to separate blood cells from the serum. The serum was diluted 1: 1 with phosphate buffer, and 0.1 ml of the solution was placed in the wells of microplates. 0.5 ml of Micrococcus lysodeikticus bacterial suspension (25 mg bacteria/100 ml phosphate buffer) (Sigma Chemical Co.) was added. Absorbance was measured directly after the addition of bacteria (E0) and after 1, 2, 3 and 30 min (final E). The final absorbance was subtracted from the initial absorbance (E0) to determine lysozyme activity with the use of a standard curve. The standard curve was plotted based on the optical density values for known lysozyme concentrations.

Nitric oxide assay

NO production was assessed by measuring nitrite accumulation in 72h culture supernatants using the Griess reaction. Briefly, 100 µl of 0.5% sulfanilamide and 0.05% N-naphtyl-ethylenediamine hydrochloride in 2.5% H3PO4 (Griess reagent) [22] were added to 100 µl of supernatants and incubated for 5 min at room temperature in the dark. The absorbance was then measured at 550 nm and nitrite concentrations were extrapolated from a sodium nitrite standard curve.

Statistical analysis

The GRAPHPAD (ISI Software, Philadelphia, PA, USA) computer program was used to conduct regression analysis and to plot collected data. Data were expressed as means ± standard error (SE). Assessment of the results was performed using one-way analysis of variance (ANOVA) procedure followed by Duncan’s Multiple Range test. The 0.05 level of probability was used as the criterion for significance [23].

RESULTS

Leukogram changes

In the present work, regarding to the result of leukogram, the total leukocytes, neutrophil, lymphocyte and monocyte were significantly decreased in CYP - immunosuppressed group when compared with the control group. In CYP- immunosuppressed animals treated with Lac in GP. (3), levamisole in GP. (6) and both Lac and levamisole in GP (7), these parameters were improved when compared with CYP treated groups. The highest improvement was found in the CYP -immunosuppressed rats treated with lactoferrin and levamisole together. In lactoferrin, levamisole treated groups, the leukogram showed nositigic change in comparison with control group (Table1).

Table 1. Changes on leukogram (TLC, neutrophils, lymphocyte, eosinophil, basophil and monocyte) in all experimental groups

<table>
<thead>
<tr>
<th>Group</th>
<th>TLC x10^3/µl</th>
<th>Neutrophils x10^3/µl</th>
<th>Lymphocyte x10^3/µl</th>
<th>Eosinophil x10^3/µl</th>
<th>Basophil x10^3/µl</th>
<th>Monocyte x10^3/µl</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I</td>
<td>8.51 ± 0.21</td>
<td>2.90 ± 0.34</td>
<td>4.30 ± 0.12</td>
<td>0.81 ± 0.34</td>
<td>0.06 ± 0.01</td>
<td>0.42 ± 0.04</td>
</tr>
<tr>
<td>Group II</td>
<td>4.45 ± 0.06</td>
<td>1.58 ± 0.08</td>
<td>2.05 ± 0.05</td>
<td>0.70 ± 0.09</td>
<td>0.00 ± 0.00</td>
<td>0.11 ± 0.02</td>
</tr>
<tr>
<td>Group III</td>
<td>6.28 ± 0.16</td>
<td>1.95 ± 0.06</td>
<td>3.23 ± 0.14</td>
<td>0.78 ± 0.09</td>
<td>0.02 ± 0.00</td>
<td>0.29 ± 0.04</td>
</tr>
<tr>
<td>Group IV</td>
<td>9.17 ± 0.14</td>
<td>3.15 ± 0.33</td>
<td>4.56 ± 0.09</td>
<td>0.89 ± 0.26</td>
<td>0.06 ± 0.01</td>
<td>0.50 ± 0.06</td>
</tr>
<tr>
<td>Group V</td>
<td>5.61 ± 0.48</td>
<td>1.69 ± 0.18</td>
<td>2.98 ± 0.25</td>
<td>0.69 ± 0.10</td>
<td>0.01 ± 0.01</td>
<td>0.21 ± 0.06</td>
</tr>
<tr>
<td>Group VI</td>
<td>8.52 ± 0.45</td>
<td>2.95 ± 0.34</td>
<td>4.14 ± 0.08</td>
<td>0.84 ± 0.38</td>
<td>0.06 ± 0.01</td>
<td>0.40 ± 0.05</td>
</tr>
<tr>
<td>Group VII</td>
<td>6.44 ± 0.38</td>
<td>2.11 ± 0.04</td>
<td>3.70 ± 0.37</td>
<td>0.81 ± 0.06</td>
<td>0.02 ± 0.00</td>
<td>0.29 ± 0.04</td>
</tr>
</tbody>
</table>

p value | <0.001 | 0.001 | <0.001 | 0.995 | 0.002 | <0.001 |

Immunological parameter changes

As shown in Tables (2, 3), significant decrease in serum total proteins, albumin, α-globulin, β-globulin and γ-globulin levels, Nitric oxide (NO) production and lysozyme activity was found in CYP -immunosuppressed group when compared with the control.
Table 2. Changes on serum total proteins, albumin, α-globulin, β-globulin and γ-globulin in all experimental groups

<table>
<thead>
<tr>
<th>Group</th>
<th>T. Protein g/dl</th>
<th>Albumin g/dl</th>
<th>α-globulin g/dl</th>
<th>β-globulin g/dl</th>
<th>γ-globulin g/dl</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I</td>
<td>8.89±0.06</td>
<td>4.03±0.05</td>
<td>1.58±0.12</td>
<td>1.41±0.13</td>
<td>1.87±0.06</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Group II</td>
<td>4.16±0.38</td>
<td>1.81±0.11</td>
<td>0.97±0.04</td>
<td>0.87±0.04</td>
<td>0.51±0.03</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Group III</td>
<td>5.87±0.40</td>
<td>2.61±0.31</td>
<td>1.17±0.09</td>
<td>1.17±0.06</td>
<td>0.92±0.03</td>
<td>0.004</td>
</tr>
<tr>
<td>Group IV</td>
<td>8.20±0.93</td>
<td>3.22±0.13</td>
<td>1.51±0.04</td>
<td>1.37±0.03</td>
<td>2.10±0.35</td>
<td>&gt;0.05</td>
</tr>
<tr>
<td>Group V</td>
<td>5.54±0.18</td>
<td>2.56±0.15</td>
<td>1.16±0.02</td>
<td>1.05±0.05</td>
<td>0.77±0.03</td>
<td>0.001</td>
</tr>
<tr>
<td>Group VI</td>
<td>7.60±0.64</td>
<td>3.06±0.12</td>
<td>1.49±0.06</td>
<td>1.28±0.16</td>
<td>1.77±0.03</td>
<td>0.001</td>
</tr>
<tr>
<td>Group VII</td>
<td>6.54±0.18</td>
<td>2.68±0.07</td>
<td>1.30±0.07</td>
<td>1.23±0.05</td>
<td>1.33±0.13</td>
<td>0.001</td>
</tr>
</tbody>
</table>

Table 3. Changes on Nitric oxide (NO) production and lysozyme activity in all experimental groups

<table>
<thead>
<tr>
<th>Group</th>
<th>NO µg/ml</th>
<th>p value</th>
<th>γ-lysozyme mg/l</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I</td>
<td>0.17±0.02</td>
<td>8.63±0.21</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group II</td>
<td>0.08±0.01</td>
<td>3.8±0.18</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group III</td>
<td>0.14±0.03</td>
<td>5.43±0.32</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group IV</td>
<td>0.18±0.02</td>
<td>7.96±0.99</td>
<td>0.086</td>
<td>0.002</td>
</tr>
<tr>
<td>Group V</td>
<td>0.14±0.02</td>
<td>5.46±0.88</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group VI</td>
<td>0.17±0.02</td>
<td>7.96±1.08</td>
<td></td>
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<tr>
<td>Group VII</td>
<td>0.16±0.03</td>
<td>7.43±0.37</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

DISCUSSION

Alkylation agents such as cyclophosphamide (CYP) were developed and introduced into clinical medicine in the 1950s. They were primarily designed as anti-cancer drugs [24]. History has shown that CYP is one of the most potent immunosuppressive drugs. In the present study, we found that CYP induced leukopenia, neutropenia and lymphopenia in rats, these results parallel to the result of [25] who demonstrated that CYP is an effective inhibitor of cell mediated immune response and leads to a depletion of lymphocytes in the peripheral blood and tissue. Lactoferrin causes an improvement in leukogram in rats with CYP immunosuppression, we agreed with [26] who said that mice treated with CYP/Lac have a higher content of functional phagocytes (neutrophils and eosinophils) and absolute cell numbers in circulation. Levamisole is an anti-inflammatory agent that also apparently enhances immune responsiveness. It is believed that levamisole mediates the immune function of T-cells and stimulates phagocytosis by monocytes [27]. Its immunostimulating effects are greater in immune-compromised animals [28]. According to [29], levamisole is a useful immunostimulatory agent in cancer patients and other patients with impaired cellular immune responses. Levamisole in our results, showed improvement of leukogram in immunosuppressed rats, we agreed with the study of [30] which focused on levamisole, and found that the lymphocyte and monocyte percentage were increased on days 7-14 after drug administration and the results of [31] which showed a significant increase in neutrophil and monocyte levels in levamisole treated animals .In addition to these findings, we found that lactoferrin modulate and improved the immunostimulatory effect of levamisole.
Cyclophosphamide exerts a direct impact on plasma cells, and it inhibits protein synthesis. As shown in Table 2, CYP group showed significant decrease of total protein, albumin, α-globulin, β-globulin compared with the control group. This result may be due to decrease protein synthesis as a result of liver damage caused by cyclophosphamide as mentioned by [32] who found that hypoproteinemia was observed in rats administered CYP (150 mg/kg) for two days. Also, we found that the CYP-immunosuppressed rats showed significant decrease in γ-globulins levels, these findings are parallel to the results observed by [13]. This may be due to the toxic effects of various chemotherapeutic on the liver, including cyclophosphamide, which was found to inhibit selected hepatic enzymes as described by [33]. CYP exerts a specific and not directly toxic effect on the cytochrome P450 system, which converts cyclophosphamide to active metabolites. As the process intensifies (following the administration of higher doses of the drug), hepatocyte dysfunction is observed, including disturbances in the synthesis of selected proteins. A decrease in the levels of γ-globulins, may be indirectly caused by the inhibition of B cell proliferation by cyclophosphamide [33]. In our study, cyclophosphamide decrease the activity of lysozyme (which is also a γ-globulin), produced by phagocytes and NO production. We agreed with Zhao et al [34] who reported a decrease in serum lysozyme levels in rats administered CYP and the study of [35] that revealed lower levels of nitrogen oxide (NO) following the administration of cyclophosphamide. The immunomodulatory effects of Lactoferrin include influences of the production and release of cytokines such as tumor necrosis factor-α [36], IL-1β [37], IL-8 [38] and nitric oxide [39]. We found that lactoferrin and levamisole elevate the levels of total protein and gamma globins levels when compared with CYP group, this agreed with the result of [13] who found that lactoferrin increased the level of total protein and γ-globulin and the result of [31] who said that Levamisole also significantly increased the total protein (7-14 days) and gamma globins levels.

In conclusion, we have demonstrated for the first time that lactoferrin have a synergistic effect to modulate and increase the immunostimulatory effect of levamisole in cyclophosphamide induced immunosuppression in rats.

Competing interests
The author declares that they have no competing interests.

REFERENCES


